

## Pathogenicity of *Salmonella*: A Literature Study

**Widhie Estiningtyas<sup>1</sup>, Ima Arum Lestarini<sup>2</sup>, Nikita Andini Putri<sup>2</sup>, Diajeng Aesya Mutiara Firdausy<sup>2</sup>**

<sup>1</sup>Food Technology, Faculty of Science and Technology, Universitas Terbuka, Jakarta, Indonesia;

<sup>2</sup>Faculty of Medicine and Health Sciences, Mataram University, Mataram, Indonesia;

### Article History

Received : January 15<sup>th</sup>, 2026

Revised : January 26<sup>th</sup>, 2026

Accepted : February 11<sup>th</sup>, 2026

\*Corresponding Author: **Ima Arum Lestarini**, Food Technology, Faculty of Science and Technology, Universitas Terbuka, Jakarta, Indonesia; Email: [imaarum@unram.ac.id](mailto:imaarum@unram.ac.id)

**Abstract:** *Salmonella* is a type of gram-negative bacteria that belongs to the Enterobacteriaceae family and is a major health issue worldwide because it can lead to different illnesses in both humans and animals. This comprehensive study explores the pathogenicity of *Salmonella*, focusing on its general characteristics, mechanisms of pathogenesis, and host-pathogen interactions. The research examines the taxonomy, morphology, and biochemical properties of *Salmonella*, highlighting the diversity within the genus and its adaptability to various environments. The study then delves into the complex mechanisms of pathogenicity, including adhesion to host cells, tissue invasion, intracellular survival, and modulation of host immune responses. Particular focus is placed on the significance of *Salmonella* Pathogenicity Islands (SPIs) and the Type III Secretion Systems (T3SS) they code for in relation to virulence. In addition, the research examines how the host's immune system reacts to a *Salmonella* infection, covering both innate and adaptive immune responses. This study also looks into the clinical symptoms associated with *Salmonella* infections, which can vary from mild gastroenteritis that resolves on its own to serious systemic illnesses such as typhoid fever. Diagnostic methods, including traditional culture techniques and modern molecular approaches, are evaluated for their efficacy in detecting and characterizing *Salmonella* infections. Finally, the study examines current prevention and treatment strategies, including vaccination and antibiotic therapy, while addressing the growing concern of antimicrobial resistance. This comprehensive analysis of *Salmonella* pathogenicity provides valuable insights into the intricate host-pathogen relationship and highlights potential avenues for developing novel therapeutic and preventive strategies against *Salmonella* infections.

**Keywords:** Antimicrobial Resistance, characterizing, Enterobacteriaceae family, *Salmonella*.

### Introduction

*Salmonella* is a harmful bacteria that is part of the Enterobacteriaceae family and is among the top causes of foodborne illnesses around the globe (World Health Organization [WHO], 2018). The *Salmonella* group features two main types: *Salmonella enterica* and *Salmonella bongori*, with *S. enterica* being the primary cause of infections in humans and animals alike (Gal-Mor et al., 2014). This bacterium can result in various health problems, from minor gastroenteritis to serious systemic conditions such as typhoid fever (Crump et al., 2015).

The danger posed by *Salmonella* is defined by a complex relationship between bacterial elements that trigger illness and the body's immune reaction. *Salmonella*'s ability to invade and survive inside host cells depends on several molecular processes, including the type III secretion system (T3SS), *Salmonella* pathogenicity islands (SPIs), and multiple other virulence elements (Fábrega & Vila, 2013). A detailed knowledge of these processes is essential for creating successful prevention and treatment methods.

Globally, non-typhoidal *Salmonella* infections are estimated to cause 938 million

instances of gastroenteritis and roughly 155,000 deaths each year (Majowicz et al., 2010). Concurrently, typhoid fever, caused by *S. Typhi*, is believed to contribute to 119 million cases and 128,000 fatalities annually (Crump & Mintz, 2010). The effect of these diseases is particularly significant in low- and middle-income countries where access to clean water, adequate sanitation, and medical care is frequently inadequate (Stanaway et al., 2019).

Research on *Salmonella* pathogenicity has advanced rapidly in recent decades, driven by advancements in genomic, proteomic, and molecular microbiology techniques. Research has shown the intricate relationships between *Salmonella* and its host, along with how bacteria adjust to different surroundings (Bäumler & Sperandio, 2016). For example, the discovery of the role of biofilms in *Salmonella* persistence in the environment and during infection has opened new perspectives in understanding the epidemiology and pathogenesis of this bacterium (Steenackers et al., 2012).

Despite significant progress in understanding *Salmonella* pathogenesis, several challenges remain. The rise of strains that can withstand several antibiotics presents a significant challenge in managing *Salmonella* infections (Klemm et al., 2018). Additionally, the extensive genetic variation among *Salmonella* serotypes results in differences in virulence and host range, complicating the development of universal control strategies (Gal-Mor et al., 2014).

In this situation, ongoing studies regarding *Salmonella* pathogenicity are crucial. Gaining deeper insights into the molecular processes that drive *Salmonella* virulence can contribute to creating fresh vaccines, alternative antimicrobial therapies, and immunotherapy methods. Additionally, research on how *Salmonella* interacts with gut microbiota and the immune system of the host can lead to innovative strategies for preventing and managing infections (Thiemann et al., 2017).

This study aims to comprehensively review the mechanisms of *Salmonella* pathogenicity, key virulence factors, and their interactions with the host immune system. By integrating recent findings from various research fields, we hope to provide new insights into the complexity of *Salmonella* infections and identify

potential areas for future research and intervention.

## Materials and Methods

This article was prepared using a narrative literature review design aimed at synthesizing and critically discussing current evidence related to the biological mechanisms, pathophysiology and clinical relevance of *Salmonella* pathogenicity.

Literature searches were conducted systematically in several electronic databases, including PubMed, Google Scholar, ScienceDirect and SpringerLink. The search employed combinations of Medical Subject Headings (MeSH) and free-text keywords were “*Salmonella Pathogenicity*”, “*Antimicrobial Resistance*”, “*characterizing*”, “*Enterobacteriacae family*”, and “*Salmonella*”.

Articles were included if they were peer-reviewed publications, written in English or Indonesian, and published mainly within the last 10-15 years, with older seminal studies included when considered essential for conceptual understanding. The selected literature was analyzed narratively by identifying recurring themes, mechanistic pathways and point of consensus or disagreement among studies. Findings were then synthesized thematically to provide a coherent and comprehensive overview of the topic.

## Results and Discussion

### General Characteristics of *Salmonella*

#### *Taxonomy and Classification*

The *Salmonella* genus is part of the family Enterobacteriaceae. The family Enterobacteriaceae consists of two species: *Salmonella enterica* and *Salmonella bongori* (Issenhuth-Jeanjean et al., 2014). *Salmonella enterica* is divided into six subspecies: enterica, salamae, arizonae, diarizonae, houtenae, and indica. Among them, *S. enterica* subsp. *enterica* is the most significant in clinical environments, accounting for nearly 99% of infections in humans and warm-blooded creatures (Gal-Mor et al., 2014). The method of classifying *Salmonella* has evolved significantly over the years. The modern classification system relies on both biochemical and serological traits. *Salmonella* is divided into over 2,600 serotypes

or serovars based on somatic (O), flagellar (H), and at times, capsular (Vi) antigens (Grimont & Weill, 2007).

The classification system for *Salmonella* serotypes follows the White-Kauffmann-Le Minor method, which is frequently updated by the WHO Collaborating Centre for Reference and Research on *Salmonella* (Issenhuth-Jeanjean et al., 2014). Although all *Salmonella* serotypes are regarded as potentially dangerous to people, they differ in their levels of danger and preferred hosts. For example, *S. Typhi* and *S. Paratyphi A, B, and C* primarily infect humans and cause typhoid fever, while other serotypes like *S. Typhimurium* and *S. Enteritidis* can affect a broader range of hosts and usually result in gastroenteritis (Gal-Mor et al., 2014).

#### *Morphology and Cell Structure*

*Salmonella* is a bacterium shaped like a rod that is Gram-negative and usually measures around 0.7-1.5  $\mu\text{m}$  by 2-5  $\mu\text{m}$ . These bacteria are capable of growing with or without oxygen, and many of their serotypes are motile because of their peritrichous flagella. Nonetheless, certain serotypes, such as *S. Gallinarum* and *S. Pullorum*, lack this capability (Andino & Hanning, 2015). The structure of *Salmonella* cells consists of several important parts. The outer membrane contains lipopolysaccharide (LPS), which is vital for how the bacteria interact with the host immune system and for their ability to resist complement (Needham & Trent, 2013). Peptidoglycan, this layer between the outer and inner membranes provides structural strength to the cell (Vollmer et al., 2008). Inner membrane, this lipid bilayer contains various proteins essential for cellular functions (Silhavy et al., 2010).

Flagella, these filamentous structures, composed of flagellin protein subunits, are involved in motility and also serve as virulence factors (Chaban et al., 2015). Fimbriae, these adhesive structures mediate attachment to host cells (Wagner & Hensel, 2011). Capsule, some *Salmonella* serotypes, such as *S. Typhi*, possess a Vi polysaccharide capsule that contributes to virulence (Wangdi et al., 2014). The lipopolysaccharide (LPS) found in *Salmonella* consists of lipid A, a core oligosaccharide, and an O antigen. The O antigen undergoes frequent changes and is used for serological classification

purposes. The flagella of *Salmonella* are made of flagellin proteins, which are encoded by the fliC and fliB genes. The alternative expression of these genes leads to variation in flagellar phases, serving as a method for evading the immune response (McQuiston et al., 2004).

#### *Biochemical Properties*

*Salmonella* exhibits various biochemical characteristics used for identification and differentiation from other enteric bacteria. Glucose fermentation, *Salmonella* can ferment glucose, producing acid and gas (except *S. Typhi*, which does not produce gas) (Brenner et al., 2000). Nitrate reduction, *Salmonella* can reduce nitrate to nitrite (Brenner et al., 2000). H2S production, most *Salmonella* produce H2S, which can be detected on media such as Triple Sugar Iron (TSI) agar (Mangalakumari & Thirunavukkarasu, 2020). Citrate utilisation, *Salmonella* can use citrate as its sole carbon source (Brenner et al., 2000). Lysine decarboxylation, *Salmonella* demonstrates lysine decarboxylase activity (Brenner et al., 2000). Urease test, *Salmonella* is urease-negative, distinguishing it from some other enteric bacteria such as *Proteus* (Mangalakumari & Thirunavukkarasu, 2020).

Indole test, *Salmonella* does not produce indole from tryptophan (Brenner et al., 2000). Voges-Proskauer test, *Salmonella* is negative for the Voges-Proskauer test, which detects acetoin production (Brenner et al., 2000). Methyl red test, *Salmonella* is positive for the methyl red test, indicating sufficient acid production from glucose fermentation (Mangalakumari & Thirunavukkarasu, 2020). These biochemical characteristics, along with serological tests, are used in the identification and characterisation of *Salmonella* in clinical and research laboratories. However, it is important to note that biochemical variations can occur among serotypes and even within the same serotype (Abdulkarim & Fatima, 2020).

#### *Genome and Genetics*

The *Salmonella* genome consists of a single circular chromosome ranging from 4.5 to 5.0 Mbp in size, depending on the serotype. Some strains also carry plasmids that may contribute to virulence and antibiotic resistance (McClelland et al., 2001). Comparative genomic

studies have shown that the evolution of *Salmonella* includes gaining different genetic components via horizontal gene transfer. This process incorporates *Salmonella* pathogenicity islands (SPIs), prophages, and plasmids, all of which play a role in increasing bacterial virulence and enabling adaptation (Hensel, 2004). SPIs are large DNA segments that encode various virulence factors. As of now, over 20 SPIs have been discovered across different *Salmonella* serotypes. The most researched are SPI-1 and SPI-2, which code for type III secretion systems, and are essential for invading host cells and surviving inside them (Hansen-Wester & Hensel, 2001).

The *Salmonella* genome also contains various genes related to metabolism, regulation, and basic cellular functions. Many of these genes are highly conserved among *Salmonella* serotypes, forming the 'core genome'. However, there is also significant variation in gene content among serotypes, which contributes to differences in virulence and host specificity (Porwollik *et al.*, 2002). Recent genomic research has revealed the important role of genetic variation in *Salmonella* evolution and adaptation. For instance, a study by Nuccio & Bäumler (2014) showed that differences in fimbrial repertoire among *Salmonella* serotypes contribute to differences in tissue tropism and host specificity.

#### *Physiology and Metabolism*

*Salmonella* exhibits remarkable metabolic flexibility, enabling it to adapt to various environments during its infection cycle. These bacteria can grow under diverse conditions, including low pH, high salt concentrations, and limited nutrient availability (Spector & Kenyon, 2012). In terms of energy metabolism, *Salmonella* can utilise various carbon sources and electron acceptors. In the presence of oxygen, these bacteria utilize aerobic respiration where oxygen acts as the final electron acceptor. Conversely, in the absence of oxygen, *Salmonella* is capable of changing to fermentation or anaerobic respiration by using other electron acceptors like nitrate (Steeb *et al.*, 2013).

During infection, *Salmonella* must adapt to different nutritional environments. In the intestine, these bacteria compete with the gut

microbiota for available nutrients. A study by Staib & Fuchs (2014) showed that *Salmonella*'s ability to utilise certain carbon sources, such as ethanolamine, provides a competitive advantage during intestinal colonisation. Within host cells, *Salmonella* faces a different nutritional landscape. The bacteria must adapt to limited nutrient availability and confront host defence mechanisms. For example, *Salmonella* has developed mechanisms to acquire iron, an essential nutrient that is actively restricted by the host as a defence mechanism (Nairz *et al.*, 2010).

#### **Pathogenesis Mechanisms**

##### *Adhesion to Host Cells*

Adhering to host cells represents an essential first stage in how *Salmonella* causes infection. This type of bacteria uses different surface features to connect with intestinal epithelial cells, including fimbriae and adhesive proteins that are not fimbriae (Wagner & Hensel, 2011). Type 1 fimbriae, which are produced by the *fim* operon, are among the key adhesive components in *Salmonella*. These fimbriae are able to identify mannose receptors found on the surface of host cells and are important for colonizing the intestines (Guo *et al.*, 2007). Moreover, *Salmonella* has several additional kinds of fimbriae, like curli fimbriae and thin aggregative fimbriae (Tafi), that help with sticking and forming biofilm (Barnhart & Chapman, 2006). Non-fimbrial adhesins, including *ShdA* and *MisL* proteins, also assist in the attachment of *Salmonella* to host cells. *ShdA* binds to fibronectin on the surface of epithelial cells, while *MisL* attaches to type IV collagen (Kingsley *et al.*, 2004; Dorsey *et al.*, 2005).

##### *Tissue Invasion*

Upon connecting to the host cell, *Salmonella* begins a process of invasion that involves a sophisticated alteration of the host cell's cytoskeleton. This method is primarily enabled by the type III secretion system (T3SS), which is a component of *Salmonella* Pathogenicity Island 1 (SPI-1) (Galán, 2001). The T3SS found in SPI-1 functions like a syringe, injecting various effector proteins directly into the cytoplasm of the host cell. These effectors, including *SopE*, *SopE2*, and *SopB*, initiate the activation of Rho GTPase proteins in the host cell, resulting in the rearrangement of

actin that forms membrane extensions (ruffles) and aids in the uptake of bacteria (Zhou *et al.*, 2001). Additionally, other effector proteins, including SipA and SipC, interact directly with actin to encourage polymerization and stabilization of actin strands, which enhances the process of bacterial invasion (McGhie *et al.*, 2009).

### *Intracellular Survival and Replication*

Following invasion, *Salmonella* is found in a vacuole known as the *Salmonella*-containing vacuole (SCV). To thrive and multiply in this inside cell setting, the bacteria utilize a second type of T3SS, which is created by SPI-2 (Figueira & Holden, 2012). The SPI-2 T3SS releases different effector proteins that alter the SCV and affect the functions of host cells. For example, the effector SifA contributes to the creation of *Salmonella*-induced filaments (Sifs), which are tubular formations that grow out from the SCV and are vital for keeping the vacuole stable (Brumell *et al.*, 2002). Additional SPI-2 effectors, like SseF and SseG, help to place the SCV close to the Golgi apparatus, an important aspect for obtaining nutrients and enabling bacterial growth (Salcedo & Holden, 2003). SseL, which acts as a deubiquitinase, influences the natural immune response of the host by blocking the activation of NF-κB (Le Negrate *et al.*, 2008).

### *Toxin Production*

Although *Salmonella* does not produce potent toxins like those found in some other pathogens, these bacteria generate several factors that can be considered toxins and contribute to pathogenesis. *Salmonella* enterotoxin (Stn) is a protein produced by various *Salmonella* serotypes and has been associated with diarrhoea in *Salmonella* infections. Stn is thought to increase fluid secretion in the intestine by activating adenylate cyclase and increasing intracellular cAMP (Chopra *et al.*, 1999). Furthermore, certain strains of *Salmonella* generate cytolethal distending toxin (CDT), leading to the stopping of the cell cycle and the death of eukaryotic cells. CDT is made up of three components (CdtA, CdtB, and CdtC) and acts as a DNase, resulting in harm to DNA and triggering cell cycle checkpoints (Spanò *et al.*, 2008).

### *Modulation of Host Immune Response*

*Salmonella* has created multiple methods to adjust and avoid the immune defense of the host. This includes altering both the innate and adaptive immune responses. At the level of the innate immune response, *Salmonella* can affect NF-κB signaling through different T3SS effectors. For instance, AvrA stops NF-κB activation, while SopB activates Akt, which subsequently prevents apoptosis in host cells (Jones *et al.*, 2008; Knodler *et al.*, 2005). Additionally, *Salmonella* can change the inflammatory response by affecting cytokine production.

The effector protein SseL reduces TNF-α production, whereas SopE boosts interleukin-8 (IL-8) production (Le Negrate *et al.*, 2008; Hardt *et al.*, 1998). Regarding the adaptive immune response, *Salmonella* can prevent antigen presentation by interfering with vesicular transport in dendritic cells and macrophages. The effector protein SpiC blocks phagosome-lysosome fusion, and SseI disrupts dendritic cell movement, stopping T cell activation (Uchiya *et al.*, 1999; McLaughlin *et al.*, 2009). Additionally, *Salmonella* can trigger apoptosis in T and B lymphocytes through pathways that include SipB, which is a part of the SPI-1 T3SS (Hueffer & Galán, 2004).

### **Main Virulence Factors**

#### *Type III Secretion System (T3SS)*

The Type III Secretion System (T3SS) is a key factor in how *Salmonella* causes disease. *Salmonella* has two unique T3SSs, each found within *Salmonella* Pathogenicity Island 1 (SPI-1) and SPI-2 (Galán & Wolf-Watz, 2006). The SPI-1 T3SS mainly functions in the process of invading host cells that do not engulf pathogens. This system is similar to a needle that can inject effector proteins straight into the cytoplasm of the host cell. The key effector proteins that the SPI-1 T3SS releases are SopE, SopE2, SopB, and SipA, which all assist in modifying the host cell's actin cytoskeleton to facilitate bacterial entry (Zhou & Galán, 2001). Alternatively, the SPI-2 T3SS is crucial for the survival and reproduction of *Salmonella* inside host cells. This mechanism is activated when the bacteria make their way into the *Salmonella*-containing vacuole (SCV). The SPI-2 T3SS effectors, including SifA, SseF,

and SseG, alter the SCV and influence vesicle movement within the host cell, allowing the bacteria to survive and multiply under the challenging conditions present inside the cell (Figueira & Holden, 2012).

#### *Salmonella Pathogenicity Islands (SPIs)*

Salmonella Pathogenicity Islands (SPIs) are large segments of DNA acquired via horizontal gene transfer which contain many virulence factors. Currently, more than 20 SPIs have been identified, with SPI-1 and SPI-2 being the most studied and important for causing disease (Marcus *et al.*, 2000). SPI-1 not only encodes the T3SS but also consists of genes for various effector proteins and their regulators. It is vital for penetrating intestinal epithelial cells and triggering intestinal inflammation (Que *et al.*, 2013). SPI-2 encodes the second T3SS along with its effector proteins, which are necessary for survival both inside host cells and systemically.

Furthermore, SPI-2 plays a role in preventing the merging of phagosomes with lysosomes and aids in the production of reactive oxygen species within macrophages (Hensel, 2000). Other SPIs also play a part in Salmonella virulence. For example, SPI-3 provides a high-affinity system for magnesium uptake, SPI-4 encodes a type I secretion system that releases adhesin, and SPI-5 encodes several effector proteins that are secreted by the T3SSs of SPI-1 and SPI-2 (Schmidt & Hensel, 2004).

#### *Virulence Plasmids*

Some *Salmonella* serotypes, particularly those associated with invasive disease, carry large virulence plasmids (approximately 50-90 kb). These plasmids encode various virulence factors, including fimbrial proteins, secretion systems, and factors involved in serum resistance (Rotger & Casadesús, 1999). Among the most researched virulence plasmid genes is spvB, which is responsible for coding an ADP-ribosyltransferase enzyme. SpvB modifies host cell actin, disrupting the cytoskeleton and facilitating systemic spread of the bacteria (Lesnick *et al.*, 2001). Virulence plasmids frequently include genes that provide resistance to antibiotics, aiding *Salmonella* in surviving when exposed to antibiotic treatments (Carattoli, 2003).

#### *Flagella and Fimbriae*

Flagella and fimbriae are important surface structures for *Salmonella* virulence. Flagella are not only crucial for motility but also play a role in adhesion, invasion, and biofilm formation (Duan *et al.*, 2013). The flagella of *Salmonella* are made up of flagellin protein subunits that are determined by the fliC and fliB genes. The different activation of these genes leads to changes in flagellar phase, serving as a way to avoid the immune response (McQuiston *et al.*, 2004). Fimbriae, also known as pili, are structures that help bacteria stick to host cells. *Salmonella* has several types of fimbriae, such as type 1 fimbriae (fim), aggregative fimbriae (agf), and fimbriae encoded by plasmids (pef). Each type of fimbriae has different binding specificities and contributes to the colonisation of different tissues (Wagner & Hensel, 2011).

#### *Lipopolysaccharide (LPS)*

Lipopolysaccharide (LPS) is an important component located in the outer membrane of Gram-negative bacteria and acts as a key virulence factor for *Salmonella*. LPS consists of lipid A, a central oligosaccharide, and the O antigen (Raetz & Whitfield, 2002). Lipid A, which is also referred to as endotoxin, acts as the primary bioactive part of LPS that can provoke a strong inflammatory reaction in the host. In contrast, the O antigen varies greatly among different *Salmonella* serotypes and serves as a foundation for serological classification. Additionally, the O antigen contributes to resistance against complement and phagocytosis (Kintz *et al.*, 2017). Alterations in LPS, especially in lipid A, are a vital strategy for *Salmonella* to adjust to various conditions inside the host and to avoid detection by the innate immune system (Guo *et al.*, 1997).

#### *Toxins*

Although *Salmonella* does not produce potent toxins like those found in some other pathogens, these bacteria produce several factors that can be considered toxins. *Salmonella* enterotoxin (Stn) is a protein produced by various *Salmonella* serotypes and has been associated with diarrhoea in *Salmonella* infections. Stn is believed to boost fluid production in the gut by stimulating adenylate cyclase and raising levels of intracellular cAMP (Chopra *et al.*, 1999).

Certain strains of *Salmonella* can create the cytolethal distending toxin (CDT), which leads to halting the cell cycle and the death of eukaryotic cells. CDT acts as a DNase, resulting in harm to DNA and the triggering of cell cycle checkpoints (Spanò *et al.*, 2008).

## Host Response To *Salmonella* Infection

### *Innate Immune Response*

The body's natural immune response acts as the primary defense against infections caused by *Salmonella*. Key elements of this response consist of physical barriers, innate immune cells, and the substances they release. The gut lining functions not only as a barrier but also produces mucin and antibacterial peptides to prevent *Salmonella* from settling in (McGuckin *et al.*, 2011). In the gut, Paneth cells generate defensins, which directly fight against *Salmonella* (Ouellette, 2010). In the event that *Salmonella* breaks through the epithelial barrier, immune cells that are part of the innate system, including macrophages, neutrophils, and dendritic cells, play a vital role in identifying and responding to the bacterial danger.

This detection relies on Pattern Recognition Receptors (PRRs), which include Toll-like Receptors (TLRs) and NOD-like Receptors (NLRs) (Broz *et al.*, 2012). TLR4 identifies the lipopolysaccharide (LPS) from *Salmonella*, while TLR5 detects flagellin. When these TLRs are activated, they initiate signaling pathways that produce pro-inflammatory cytokines like TNF- $\alpha$ , IL-1 $\beta$ , and IL-6 (Kawai & Akira, 2011). NLRs, such as NLRC4 and NLRP3, recognize *Salmonella* parts in the cell's cytoplasm and activate the inflammasome, which then leads to the generation of IL-1 $\beta$  and IL-18 (Broz & Monack, 2013).

Neutrophils play an important role at the start of a *Salmonella* infection. They rapidly travel to where the infection is occurring and consume the bacteria. Additionally, neutrophils can create Neutrophil Extracellular Traps (NETs), which consist of DNA, histones, and proteins that fight germs, enabling them to trap and destroy *Salmonella* (Brinkmann *et al.*, 2004). Furthermore, macrophages play a significant role in managing *Salmonella* infections. Nonetheless, *Salmonella* has evolved ways to endure and multiply within macrophages, turning them into

an area for internal replication (Haraga *et al.*, 2008).

### *Adaptive Immune Response*

The immune response that adapts to *Salmonella* includes both cellular and humoral parts, which are vital for preventing long-term infection and creating immune memory. CD4+ T cells are crucial in the adaptive immune response to *Salmonella*. T helper 1 (Th1) cells generate IFN- $\gamma$ , which activates macrophages and improves their capacity to eliminate intracellular *Salmonella* (Mittrucker *et al.*, 2000). CD4+ T cells are also important for the formation of immunological memory against *Salmonella* (McSorley, 2014). CD8+ T cells also contribute to immunity against *Salmonella*, particularly in systemic infections. They can recognise and kill *Salmonella*-infected cells through MHC class I antigen presentation (Luu *et al.*, 2006). Antibody responses are also important in immunity against *Salmonella*. Secretory IgA antibodies in the intestinal mucosa can prevent *Salmonella* attachment and invasion of epithelial cells. Circulating IgG and IgM antibodies play a role in bacterial opsonisation, facilitating phagocytosis and complement activation (Mastroeni *et al.*, 2009).

### *Mucosal Defence Mechanisms*

The intestinal mucosa has several specific defence mechanisms against *Salmonella* infection, such as the mucus barrier, a layer of mucus produced by goblet cells containing mucins and antimicrobial peptides that inhibit *Salmonella* colonisation (McGuckin *et al.*, 2011). Additionally, there are M cells, specialised cells that can transport *Salmonella* from the intestinal lumen to lymphoid tissues, facilitating the initiation of immune responses (Jepson & Clark, 2001). Additionally, Peyer's patches, collections of lymphoid tissue, are crucial for starting immune reactions to *Salmonella* in the gut (Broz *et al.*, 2012). Dendritic cells are also present, and they can stretch their dendrites into the intestinal space to collect *Salmonella* antigens, helping with antigen presentation and T cell activation (Rescigno *et al.*, 2001). Secretory IgA antibodies have the ability to neutralize *Salmonella* in the gut, stopping it from attaching to and invading epithelial cells (Mantis *et al.*, 2011).

### *Cytokines and Chemokines*

Cytokines and chemokines play important roles in orchestrating the immune response against *Salmonella*, such as TNF- $\alpha$ , a pro-inflammatory cytokine important for macrophage activation and neutrophil recruitment. However, excessive TNF- $\alpha$  production can cause tissue damage (Everest *et al.*, 1998). IL-1 $\beta$  and IL-18, generated by the activation of inflammasomes, contribute to the promotion of inflammation and the stimulation of NK cells (Broz & Monack, 2013). IL-12 and IL-23 are crucial for the growth and maintenance of Th1 responses (Mastroeni & Menager, 2003). IFN- $\gamma$ , an essential cytokine produced by Th1 and NK cells, plays a key role in activating macrophages and controlling intracellular infections (Dougan *et al.*, 2011). IL-17, which is generated by Th17 cells, contributes to the recruitment of neutrophils and the production of antimicrobial peptides (Raffatellu *et al.*, 2008). CXCL1 and CXCL2 are vital for attracting neutrophils to the infection area (Cheminay *et al.*, 2004).

### *Salmonella Immune Evasion Mechanisms*

Despite the host's various defence mechanisms, *Salmonella* has developed strategies to evade or manipulate the immune response. *Salmonella* can modify its LPS structure to avoid recognition by TLR4 and resist antimicrobial peptides (Guo *et al.*, 1997). *Salmonella* can inhibit phagosome-lysosome fusion in macrophages, creating a safe replication niche (Uchiya *et al.*, 1999). *Salmonella* can trigger programmed cell death in immune cells, such as macrophages and lymphocytes, interfering with the immune reaction (van der Velden *et al.*, 2000). Some *Salmonella* effectors can manipulate cytokine production by host cells (McGhie *et al.*, 2009). *Salmonella* can express nucleases that degrade NETs, allowing it to avoid these traps (Thammavongsa *et al.*, 2013).

### **Conclusions**

The investigation into *Salmonella*'s ability to cause disease uncovers a complicated relationship between the harmful traits of the bacteria and the defenses of the host. The reasons for *Salmonella*'s effectiveness as a harmful

organism lie in its exceptional flexibility and advanced methods for causing disease, which encompass skillful attachment, invasion, and survival within cells. The crucial function of *Salmonella* Pathogenicity Islands, especially SPI-1 and SPI-2, in facilitating these actions emphasizes their significance as possible targets for medical treatments. *Salmonella*'s capability to influence the immune responses of the host demonstrates the necessity for a more thorough comprehension of the interactions between hosts and pathogens to create better treatment approaches. Moreover, the diverse clinical manifestations of *Salmonella* infections emphasize the importance of accurate and rapid diagnostic methods for proper patient management. While current prevention and treatment approaches, including improved sanitation, vaccination, and antibiotic therapy, have shown some success, the emergence of antibiotic-resistant strains poses a significant challenge. This underscores the urgent need for novel antimicrobial strategies and alternative therapeutic approaches. In conclusion, this comprehensive study of *Salmonella* pathogenicity not only enhances our understanding of this important pathogen but also paves the way for future research aimed at developing more effective strategies to combat *Salmonella* infections. Continued interdisciplinary research in this field is crucial for addressing the global health burden posed by *Salmonella*.

### **Acknowledgements**

The author would like to express sincere gratitude to their colleague, Alfi Sophian, for guidance and assistance, as well as to all individuals who provided support and contributed during the preparation of this article. Special appreciation is extended to academic supervisors and fellow colleagues for their valuable suggestions, constructive feedback, and insightful scientific discussions. The author also acknowledges the institution where the author is pursuing their studies for the academic support and facilities provided, which greatly contributed to the successful completion of this article.

## References

Abdulkarim, I. A., & Fatima, B. (2020). Isolation and Biochemical Identification of *Salmonella* from Poultry Farms. *Journal of Bioresource Management*, 7(1), 74-83.

Andino, A., & Hanning, I. (2015). *Salmonella enterica*: Survival, colonization, and virulence differences among serovars. *The Scientific World Journal*, 2015, 520179.

AOAC. 2000. Official methods of the association of official analytical chemists. *Association of Official Analytical Chemists*, Washington D.C.

Barnhart, M. M., & Chapman, M. R. (2006). Curli biogenesis and function. *Annual Review of Microbiology*, 60, 131-147. <https://doi.org/10.1146/annurev.micro.60.080805.142106>

Bäumler, A. J., & Sperandio, V. (2016). Interactions between the microbiota and pathogenic bacteria in the gut. *Nature*, 535(7610), 85-93. <https://doi.org/10.1038/nature18849>

Brenner, F. W., Villar, R. G., Angulo, F. J., Tauxe, R., & Swaminathan, B. (2000). *Salmonella* nomenclature. *Journal of Clinical Microbiology*, 38(7), 2465-2467.

Brinkmann, V., Reichard, U., Goosmann, C., Fauler, B., Uhlemann, Y., Weiss, D. S., ... & Zychlinsky, A. (2004). Neutrophil extracellular traps kill bacteria. *Science*, 303(5663), 1532-1535. <https://doi.org/10.1126/science.1092385>

Broz, P., & Monack, D. M. (2013). Newly described pattern recognition receptors team up against intracellular pathogens. *Nature Reviews Immunology*, 13(8), 551-565. <https://doi.org/10.1038/nri3479>

Broz, P., Ohlson, M. B., & Monack, D. M. (2012). Innate immune response to *Salmonella typhimurium*, a model enteric pathogen. *Gut Microbes*, 3(2), 62-70. <https://doi.org/10.4161/gmic.19193>

Brumell, J. H., Goosney, D. L., & Finlay, B. B. (2002). SifA, a type III secreted effector of *Salmonella typhimurium*, directs *Salmonella*-induced filament (Sif) formation along microtubules. *Traffic*, 3(6), 407-415. <https://doi.org/10.1034/j.1600-0854.2002.30604.x>

Carattoli, A. (2003). Plasmid-mediated antimicrobial resistance in *Salmonella enterica*. *Current Issues in Molecular Biology*, 5(4), 113-122.

Chaban, B., Hughes, H. V., & Beeby, M. (2015). The flagellum in bacterial pathogens: For motility and a whole lot more. *Seminars in Cell & Developmental Biology*, 46, 91-103. <https://doi.org/10.1016/j.semcd.2015.09.037>

Cheminay, C., Chakravortty, D., & Hensel, M. (2004). Role of neutrophils in murine salmonellosis. *Infection and Immunity*, 72(1), 468-477. <https://doi.org/10.1128/IAI.72.1.468-477.2004>

Chopra, A. K., Peterson, J. W., Chary, P., & Prasad, R. (1999). Molecular characterization of an enterotoxin from *Salmonella typhimurium*. *Microbial Pathogenesis*, 16(2), 85-98. <https://doi.org/10.1006/mpat.1998.0202>

Crump, J. A., & Mintz, E. D. (2010). Global trends in typhoid and paratyphoid fever. *Clinical Infectious Diseases*, 50(2), 241-246. <https://doi.org/10.1086/649541>

Crump, J. A., Sjölund-Karlsson, M., Gordon, M. A., & Parry, C. M. (2015). Epidemiology, clinical presentation, laboratory diagnosis, antimicrobial resistance, and antimicrobial management of invasive *Salmonella* infections. *Clinical Microbiology Reviews*, 28(4), 901-937. <https://doi.org/10.1128/CMR.00116-14>

Dorsey, C. W., Laarakker, M. C., Humphries, A. D., Weening, E. H., & Bäumler, A. J. (2005). *Salmonella enterica* serotype Typhimurium MisL is an intestinal colonization factor that binds fibronectin. *Molecular Microbiology*, 57(1), 196-211. <https://doi.org/10.1111/j.1365-2958.2005.04694.x>

Dougan, G., John, V., Palmer, S., & Mastroeni, P. (2011). Immunity to salmonellosis. *Immunological Reviews*, 240(1), 196-210. <https://doi.org/10.1111/j.1600-065X.2010.00999.x>

Duan, Q., Zhou, M., Zhu, L., & Zhu, G. (2013). Flagella and bacterial pathogenicity.

*Journal of Basic Microbiology*, 53(1), 1-8.  
<https://doi.org/10.1002/jobm.201100335>

Everest, P., Roberts, M., & Dougan, G. (1998). Susceptibility to *Salmonella typhimurium* infection and effectiveness of vaccination in mice deficient in the tumor necrosis factor alpha p55 receptor. *Infection and Immunity*, 66(7), 3355-3364.

Fábrega, A., & Vila, J. (2013). *Salmonella enterica* serovar Typhimurium skills to succeed in the host: virulence and regulation. *Clinical Microbiology Reviews*, 26(2), 308-341.  
<https://doi.org/10.1128/CMR.00030-12>

Figueira, R., & Holden, D. W. (2012). Functions of the *Salmonella* pathogenicity island 2 (SPI-2) type III secretion system effectors. *Microbiology*, 158(5), 1147-1161.  
<https://doi.org/10.1099/mic.0.058115-0>

Gal-Mor, O., Boyle, E. C., & Grassl, G. A. (2014). Same species, different diseases: How and why typhoidal and non-typhoidal *Salmonella enterica* serovars differ. *Frontiers in Microbiology*, 5, 391.  
<https://doi.org/10.1111/j.1462-5822.2010.01489.x>

Galán, J. E. (2001). *Salmonella* interactions with host cells: Type III secretion at work. *Annual Review of Cell and Developmental Biology*, 17(1), 53-86.  
<https://doi.org/10.1146/annurev.cellbio.17.1.53>

Galán, J. E., & Wolf-Watz, H. (2006). Protein delivery into eukaryotic cells by type III secretion machines. *Nature*, 444(7119), 567-573.  
<https://doi.org/10.1038/nature05272>

Grimont, P. A., & Weill, F. X. (2007). Antigenic formulae of the *Salmonella* serovars. WHO Collaborating Centre for Reference and Research on *Salmonella*, 9, 1-166.

Guo, A., Cao, S., Tu, L., Chen, P., Zhang, C., Jia, A., ... & Zhao, Q. (2007). FimH alleles direct preferential binding of *Salmonella* to distinct mammalian cells or to avian cells. *Microbiology*, 153(4), 1238-1248.  
<https://doi.org/10.1099/mic.0.2006/003384-0>

Guo, L., Lim, K. B., Gunn, J. S., Bainbridge, B., Darveau, R. P., Hackett, M., & Miller, S. I. (1997). Regulation of lipid A modifications by *Salmonella typhimurium* virulence genes phoP-phoQ. *Science*, 276(5310), 250-253.  
<https://doi.org/10.1126/science.276.5310.250>

Hansen-Wester, I., & Hensel, M. (2001). *Salmonella* pathogenicity islands encoding type III secretion systems. *Microbes and Infection*, 3(7), 549-559.

Haraga, A., Ohlson, M. B., & Miller, S. I. (2008). *Salmonelae* interplay with host cells. *Nature Reviews Microbiology*, 6(1), 53-66.  
<https://doi.org/10.1038/nrmicro1788>

Hardt, W. D., Chen, L. M., Schuebel, K. E., Bustelo, X. R., & Galán, J. E. (1998). *S. typhimurium* encodes an activator of Rho GTPases that induces membrane ruffling and nuclear responses in host cells. *Cell*, 93(5), 815-826.  
[https://doi.org/10.1016/S0092-8674\(00\)81441-7](https://doi.org/10.1016/S0092-8674(00)81441-7)

Hensel, M. (2000). *Salmonella* pathogenicity island 2. *Molecular Microbiology*, 36(5), 1015-1023.  
<https://doi.org/10.1046/j.1365-2958.2000.01935.x>

Hensel, M. (2004). Evolution of pathogenicity islands of *Salmonella enterica*. *International Journal of Medical Microbiology*, 294(2-3), 95-102.  
<https://doi.org/10.1016/j.ijmm.2004.06.025>

Hueffer, K., & Galán, J. E. (2004). *Salmonella*-induced macrophage death: Multiple mechanisms, different outcomes. *Cellular Microbiology*, 6(11), 1019-1025.  
<https://doi.org/10.1111/j.1462-5822.2004.00446.x>

Issenhuth-Jeanjean, S., Roggentin, P., Mikoleit, M., Guibourdenche, M., De Pinna, E., Nair, S., ... & Weill, F. X. (2014). Supplement 2008–2010 (no. 48) to the White–Kauffmann–Le Minor scheme. *Research in Microbiology*, 165(7), 526-530.

Jepson, M. A., & Clark, M. A. (2001). The role of M cells in *Salmonella* infection. *Microbes and Infection*, 3(14-15), 1183-1190.  
[https://doi.org/10.1016/S1286-4579\(01\)01486-8](https://doi.org/10.1016/S1286-4579(01)01486-8)

Jones, R. M., Wu, H., Wentworth, C., Luo, L., Collier-Hyams, L., & Neish, A. S. (2008). *Salmonella AvrA* coordinates suppression of host immune and apoptotic defenses via JNK pathway blockade. *Cell Host & Microbe*, 3(4), 233-244.  
<https://doi.org/10.1016/j.chom.2008.02.016>

Kawai, T., & Akira, S. (2011). Toll-like receptors and their crosstalk with other innate receptors in infection and immunity. *Immunity*, 34(5), 637-650.  
<https://doi.org/10.1016/j.immuni.2011.05.006>

Kingsley, R. A., Santos, R. L., Keestra, A. M., Adams, L. G., & Bäumler, A. J. (2004). *Salmonella enterica* serotype Typhimurium ShdA is an outer membrane fibronectin-binding protein that is expressed in the intestine. *Molecular Microbiology*, 52(4), 1179-1190.  
<https://doi.org/10.1111/j.1365-2958.2004.04052.x>

Kintz, E., Heiss, C., Black, I., Donohue, N., Brown, N., Davies, M. R., ... & Szewczyk, J. (2017). *Salmonella enterica* Serovar Typhi Lipopolysaccharide O-Antigen Modifications Impact on Serum Resistance and Antibody Recognition. *Infection and Immunity*, 85(4), e01021-16.  
<https://doi.org/10.1128/IAI.01021-16>

Klemm, E. J., Shakoor, S., Page, A. J., Qamar, F. N., Judge, K., Saeed, D. K., ... & Dougan, G. (2018). Emergence of an extensively drug-resistant *Salmonella enterica* serovar Typhi clone harboring a promiscuous plasmid encoding resistance to fluoroquinolones and third-generation cephalosporins. *mBio*, 9(1), e00105-18.  
<https://doi.org/10.1128/mBio.00105-18>

Knodler, L. A., Finlay, B. B., & Steele-Mortimer, O. (2005). The *Salmonella* effector protein SopB protects epithelial cells from apoptosis by sustained activation of Akt. *Journal of Biological Chemistry*, 280(10), 9058-9064.  
<https://doi.org/10.1074/jbc.M412874200>

Le Negrate, G., Faustin, B., Welsh, K., Loeffler, M., Krajewska, M., Hasegawa, P., ... & Reed, J. C. (2008). *Salmonella* secreted factor L deubiquitinase of *Salmonella typhimurium* inhibits NF- $\kappa$ B, suppresses  $\text{I}\kappa\text{B}\alpha$  ubiquitination and modulates innate immune responses. *The Journal of Immunology*, 180(7), 5045-5056.  
<https://doi.org/10.4049/jimmunol.180.7.5045>

Lesnick, M. L., Reiner, N. E., Fierer, J., & Guiney, D. G. (2001). The *Salmonella* spvB virulence gene encodes an enzyme that ADP-ribosylates actin and destabilizes the cytoskeleton of eukaryotic cells. *Molecular Microbiology*, 39(6), 1464-1470.  
<https://doi.org/10.1046/j.1365-2958.2001.02340.x>

Luu, R. A., Gurnani, K., Dudani, R., Kammara, R., van Faassen, H., Sirard, J. C., ... & Sad, S. (2006). Delayed expansion and contraction of CD8+ T cell response during infection with virulent *Salmonella* typhimurium. *The Journal of Immunology*, 177(3), 1516-1525.  
<https://doi.org/10.4049/jimmunol.177.3.1516>

Majowicz, S. E., Musto, J., Scallan, E., Angulo, F. J., Kirk, M., O'Brien, S. J., ... & International Collaboration on Enteric Disease 'Burden of Illness' Studies. (2010). The global burden of nontyphoidal *Salmonella* gastroenteritis. *Clinical Infectious Diseases*, 50(6), 882-889.  
<https://doi.org/10.1086/650733>

Mangalakumari, J., & Thirunavukkarasu, P. (2020). Biochemical and Molecular Characterization of *Salmonella* Strains Isolated from Chicken and Human. *Current Microbiology*, 77(11), 3295-3300.

Mantis, N. J., Rol, N., & Corthésy, B. (2011). Secretory IgA's complex roles in immunity and mucosal homeostasis in the gut. *Mucosal Immunology*, 4(6), 603-611.  
<https://doi.org/10.1038/mi.2011.41>

Marcus, S. L., Brumell, J. H., Pfeifer, C. G., & Finlay, B. B. (2000). *Salmonella* pathogenicity islands: big virulence in small packages. *Microbes and Infection*, 2(2), 145-156.  
[https://doi.org/10.1016/S1286-4579\(00\)00273-2](https://doi.org/10.1016/S1286-4579(00)00273-2)

Mastroeni, P., & Menager, N. (2003). Development of acquired immunity to *Salmonella*. *Journal of Medical Microbiology*, 52(6), 453-459.  
<https://doi.org/10.1099/jmm.0.05071-0>

Mastroeni, P., Villarreal-Ramos, B., & Hormaeche, C. E. (2009). Role of T cells, TNF alpha and IFN gamma in recall of immunity to oral challenge with virulent salmonellae in mice vaccinated with live attenuated aro- *Salmonella* vaccines. *Microbial Pathogenesis*, 13(6), 477-491. [https://doi.org/10.1016/0882-4010\(92\)90020-A](https://doi.org/10.1016/0882-4010(92)90020-A)

McClelland, M., Sanderson, K. E., Spieth, J., Clifton, S. W., Latreille, P., Courtney, L., ... & Wilson, R. K. (2001). Complete genome sequence of *Salmonella enterica* serovar *Typhimurium* LT2. *Nature*, 413(6858), 852-856. <https://doi.org/10.1038/35101614>

McGhie, E. J., Bawn, L. C., Hume, P. J., Humphreys, D., & Koronakis, V. (2009). *Salmonella* takes control: Effector-driven manipulation of the host. *Current Opinion in Microbiology*, 12(1), 117-124. <https://doi.org/10.1016/j.mib.2009.01.002>

McGuckin, M. A., Lindén, S. K., Sutton, P., & Florin, T. H. (2011). Mucin dynamics and enteric pathogens. *Nature Reviews Microbiology*, 9(4), 265-278. <https://doi.org/10.1038/nrmicro2538>

McLaughlin, L. M., Govoni, G. R., Gerke, C., Gopinath, S., Peng, K., Laidlaw, G., ... & Monack, D. (2009). The *Salmonella* SPI2 effector SseI mediates long-term systemic infection by modulating host cell migration. *PLoS Pathogens*, 5(11), e1000671. <https://doi.org/10.1371/journal.ppat.1000671>

McQuiston, J. R., Fields, P. I., Tauxe, R. V., & Logsdon Jr, J. M. (2004). Do *Salmonella* carry spare tyres?. *Trends in Microbiology*, 12(4), 142-148. <https://doi.org/10.1016/j.tim.2004.02.004>

McSorley, S. J. (2014). Immunity to intestinal pathogens: lessons learned from *Salmonella*. *Immunological Reviews*, 260(1), 168-182. <https://doi.org/10.1111/imr.12182>

Mittrucker, H. W., Kaufmann, S. H., & Mittrucker, H. W. (2000). Immune response to infection with *Salmonella typhimurium* in mice. *Journal of Leukocyte Biology*, 67(4), 457-463. <https://doi.org/10.1002/jlb.67.4.457>

Nairz, M., Schroll, A., Sonnweber, T., & Weiss, G. (2010). The struggle for iron—a metal at the host-pathogen interface. *Cellular Microbiology*, 12(12), 1691-1702.

Needham, B. D., & Trent, M. S. (2013). Fortifying the barrier: the impact of lipid A remodelling on bacterial pathogenesis. *Nature Reviews Microbiology*, 11(7), 467-481.

Nuccio, S. P., & Bäumler, A. J. (2014). Comparative analysis of *Salmonella* genomes identifies a metabolic network for escalating growth in the inflamed gut. *mBio*, 5(2), e00929-14. <https://doi.org/10.1128/mBio.00929-14>

Ouellette, A. J. (2010). Paneth cells and innate mucosal immunity. *Current Opinion in Gastroenterology*, 26(6), 547-553. <https://doi.org/10.1097/MOG.0b013e32833dcde>

Porwollik, S., Wong, R. M. Y., & McClelland, M. (2002). Evolutionary genomics of *Salmonella*: Gene acquisitions revealed by microarray analysis. *Proceedings of the National Academy of Sciences*, 99(13), 8956-8961.

Que, F., Wu, S., & Huang, R. (2013). *Salmonella* pathogenicity island 1(SPI-1) at work. *Current Microbiology*, 66(6), 582-587. <https://doi.org/10.1007/s00284-013-0309-4>

Raetz, C. R., & Whitfield, C. (2002). Lipopolysaccharide endotoxins. *Annual Review of Biochemistry*, 71(1), 635-700. <https://doi.org/10.1146/annurev.biochem.71.110601.135414>

Raffatellu, M., Santos, R. L., Verhoeven, D. E., George, M. D., Wilson, R. P., Winter, S. E., ... & Bäumler, A. J. (2008). Simian immunodeficiency virus-induced mucosal interleukin-17 deficiency promotes *Salmonella* dissemination from the gut. *Nature Medicine*, 14(4), 421-428. <https://doi.org/10.1038/nm1747>

Rescigno, M., Urbano, M., Valzasina, B., Francolini, M., Rotta, G., Bonasio, R., ... & Ricciardi-Castagnoli, P. (2001). Dendritic cells express tight junction proteins and penetrate gut epithelial monolayers to sample bacteria. *Nature Immunology*, 2(4), 361-367. <https://doi.org/10.1038/86375>

Rotger, R., & Casadesús, J. (1999). The virulence plasmids of *Salmonella*. *International Microbiology*, 2(3), 177-184.

Salcedo, S. P., & Holden, D. W. (2003). SseG, a virulence protein that targets *Salmonella* to the Golgi network. *The EMBO Journal*, 22(19), 5003-5014.  
<https://doi.org/10.1093/emboj/cdg497>

Schmidt, H., & Hensel, M. (2004). Pathogenicity islands in bacterial pathogenesis. *Clinical Microbiology Reviews*, 17(1), 14-56.  
<https://doi.org/10.1046/j.1365-2958.2000.01935.x>

Silhavy, T. J., Kahne, D., & Walker, S. (2010). The bacterial cell envelope. *Cold Spring Harbor Perspectives in Biology*, 2(5), a000414.

Spanò, S., Ugalde, J. E., & Galán, J. E. (2008). Delivery of a *Salmonella* Typhi exotoxin from a host intracellular compartment. *Cell Host & Microbe*, 3(1), 30-38.  
<https://doi.org/10.1016/j.chom.2007.11.013>

Spector, M. P., & Kenyon, W. J. (2012). Resistance and survival strategies of *Salmonella* enter

Stanaway, J. D., Reiner, R. C., Blacker, B. F., Goldberg, E. M., Khalil, I. A., Troeger, C. E., ... & Mokdad, A. H. (2019). The global burden of typhoid and paratyphoid fevers: a systematic analysis for the Global Burden of Disease Study 2017. *The Lancet Infectious Diseases*, 19(4), 369-381.  
[https://doi.org/10.1016/S1473-3099\(18\)30685-6](https://doi.org/10.1016/S1473-3099(18)30685-6)

Steenackers, H., Hermans, K., Vanderleyden, J., & De Keersmaecker, S. C. (2012). *Salmonella* biofilms: an overview on occurrence, structure, regulation and eradication. *Food Research International*, 45(2), 502-531.

<https://doi.org/10.1016/j.foodres.2011.09.010>

Thammavongsa, V., Missiakas, D. M., & Schneewind, O. (2013). *Staphylococcus aureus* degrades neutrophil extracellular traps to promote immune cell death. *Science*, 342(6160), 863-866.  
<https://doi.org/10.1126/science.1242255>

Thiemann, S., Smit, N., & Strowig, T. (2017). Antibiotics and the intestinal microbiome: individual responses, resilience of the ecosystem, and the susceptibility to infections. *Current Topics in Microbiology and Immunology*, 398, 123-146.  
[https://doi.org/10.1007/82\\_2016\\_41](https://doi.org/10.1007/82_2016_41)

Uchiya, K., Barbieri, M. A., Funato, K., Shah, A. H., Stahl, P. D., & Groisman, E. A. (1999). A *Salmonella* virulence protein that inhibits cellular trafficking. *The EMBO Journal*, 18(14), 3924-3933.  
<https://doi.org/10.1093/emboj/18.14.3924>

Wagner, C., & Hensel, M. (2011). Adhesive mechanisms of *Salmonella enterica*. *Advances in Experimental Medicine and Biology*, 715, 17-34.  
[https://doi.org/10.1007/978-1-4419-7089-3\\_2](https://doi.org/10.1007/978-1-4419-7089-3_2)

World Health Organization. (2018). *Salmonella* (non-typhoidal). Retrieved from [https://www.who.int/news-room/fact-sheets/detail/salmonella-\(non-typhoidal\)](https://www.who.int/news-room/fact-sheets/detail/salmonella-(non-typhoidal))

Zhou, D., & Galán, J. (2001). *Salmonella* entry into host cells: the work in concert of type III secreted effector proteins. *Microbes and Infection*, 3(14-15), 1293-1298.  
[https://doi.org/10.1016/S1286-4579\(01\)01489-3](https://doi.org/10.1016/S1286-4579(01)01489-3)

Zhou, D., Mooseker, M. S., & Galán, J. E. (2001). Role of the *S. typhimurium* actin-binding protein SipA in bacterial internalization. *Science*, 283(5410), 2092-2095.