

## Specialty Dihydrobenzoxanthone's *Artocarpus* Purified By Vacuum Liquid Chromatography (VLC)

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**Abstract:** One family of plants that are source of bioactive chemicals is the Moraceae. *Artocarpus* is the main genus of the Moraceae. Several species of the genus *Artocarpus* have been isolated their secondary metabolites. The main fractions obtained from the VLC are analyzed again by TLC. Fractions that have same spots (Rf) pooled. Purification process on main fractions are done repeatedly by radial chromatography. Flavonoid is the most found from *Artocarpus* plant. Dihydrobenzoxanthone is one of flavonoid derivatives which is successfully isolated from *Artocarpus*. Dihydrobenzoxanthone is only formed from the flavone with ring B which is oxygenated with pattern of 2', 4' and 5'. Students can be learned dihydrobenzoxanthone's *Artocarpus* by laboratory activities.

**Keywords:** *Artocarpus*, dihydrobenzoxanthone, students.

### Introduction

Plants contain secondary metabolites that can be extracted for scientific, technological and commercial purposes. Most plant species grow in tropical climates. Indonesia as a tropical country has a very high diversity of plants. Indonesia has 25,000 high-level plant species and 40% are Indonesian endemic plants (Achmad, 1995). However, the abundant natural wealth has not been studied, recorded and reviewed, so that it cannot be utilized optimally.

In the field of traditional medicine there are many species that have reported, but research on the useful of chemicals content in plant has not much in Indonesia. One group of plants that has benefits in the field of medicine is the family of Moraceae, specifically the genus *Artocarpus* (Heyne, 1987).

*Artocarpus* belongs to the clan of jackfruit plants. Plant species included in this genus such as jackfruit or breadfruit which are widely used for fruit, wood, skin, and sap. Research on the chemical content of the genus *Artocarpus* has been carried out. The chemical compounds reported in the *Artocarpus* include triterpenes, steroids, flavonoids, stilbenoids, and lignans (Ahmad, 1996) (Nomura, 1998). The phenolic compound which is the most abundant

compound found in the genus *Artocarpus* is a class of flavonoids. These phenolic compounds are reported to have anti-bacterial activity (Khan, 2003), anti-fungal (Jayasinghe, et. al, 2004), anti-malaria (Widyawaruyanti, et. al, 2007) and cytotoxic (Syah, et. al, 2006).

Antimalarial activity-guided study of the aerial parts of *Artocarpus integer* led to the isolation of the prenylated stilbene, *trans*-4-(3-methyl-*E*-but-1-enyl)-3,5,2',4'-tetrahydroxystilbene (Boonlaksiri, 2000). Phytochemical testing of jackfruit leaf (*Artocarpus heterophyllus* Lmk) showed that belong to flavonoid compounds and antibacterial testing of flavonoid compounds *Staphylococcus aureus* with a concentration of 10.000 ppm inhibited 10.50 (Darmawati, 2015). Other research showed *Artocarpus altilis* (Parkinson) ethyl acetate extract showed cytoprotective activities (Wang, 2006).

The benefits and uses of *Artocarpus* in the health sector and easily found everywhere are an interesting aspect. This study aims to facilitate the isolation of active compounds from *Artocarpus* as basic ingredients and standards in various scientific studies such as microbiology and pharmacy. Isolated compounds can be further tested as active natural substances against various types of diseases.

## Materials and Method

### Extraction, Isolation, and Purification

The method for making extracts consists of powder, extraction, solvent separation, and extract concentration. The extraction process is repeated until the less colorful supernatant. Solvent separation used a rotary evaporator and concentrated in a water bath, so a crude extract produced.

Futhermore, the obtained extraction is underwent TLC (Thin Layer Chromatography) treatment using various eluents. TLC chromatogram is used as a basis for conducting fractionation by vacuum liquid chromatography (VLC). The main fractions obtained from the VLC are analyzed again by TLC. Fractions that have same spots (Rf) pooled. Purification process on main factions are done repeatedly by radial chromatography. TLC chromatogram is used to test the purity of an isolate, the pure compound must show a single spot on three different eluent systems. The purity test can also be done by measuring its melting point.

## Result and Discussion

### Structural Determination

The structures of pure compound are determined by spectroscopic methods: (i) UV-Vis spectrum to determine the presence of double bond conjugation in the structure of pure compound. (ii) Infrared spectrum to know the functional groups. (iii) NMR (Nuclear Magnetic Resonances) is a great tools for determinining of pure compound (Okunlola, 2019).

### Flavonoids

The flavonoid pigments, one of the most numerous and widespread groups of natural constituents, are of importance and interest not only because of their significant natural functions in the economy of the plant, but also because certain members of the group are physiologically active in humans (Harborne et al. 1991). The flavonoids are derived from the flavan or isoflavan skeleton and comprise a large

group of secondary metabolites from higher plants. The sites of plant flavonoid biosynthesis, storage and final function often differ at the subcellular, cell, and even tissue and organ levels. Efficient transport systems for flavonoids across endomembranes and the plasma membrane are therefore required (Zhao, *et al.*, 2010).

The flavonoid compounds of *Artocapus* have variety of frameworks such as chalkon derivatives, flavanones, flavan-3ol, simple flavone, prenylflavone, oxepinoflavone, pyranoflavone, dihydrobenzoxanthone, furanodihydrobenzoxanthone, pyranodihydrobenzoxanthone, quinonoxanthone, cyclopentenoxanthone, xanthonolide, dihydroxanthone, and cyclopentenoxanthone. This article will be described dihydrobenzoxanthone that have been isolated from *Artocapus*.

### Dihydrobenzoxanthone

Dihydrobenzoxanthone is formed from C6' in ring B bound directly to the carbon from the group of prenyl hexagon forms a ring. Dihydrobenzoxanthone formed from the flavone with B ring which was oxygenated with pattern of 2', 4' and 5'. The two hydroxy groups at C2' and C5' activated C6' which was located at the ortho position of hydroxy group (Hakim, 2010).

Class of dihydroxanthone compounds which have been isolated are artobiloxanthone (1) which are isolated from the stem bark of *A. scortechinii* by Ferlinahayati (Ferlinahayati, 1999) (Pratap, 2014). Compound (1) is also isolated from *A. nobilis* (Sultanbawa, et. al, 1989) (Jayasinghe ULB, et. al, 2008)). Syah et al. (2002) is succeeded in isolating artoindonesianin S (2) and artoindonesianin T (3) of stem wood of *A. champeden* (Syah, et. al, 2002). A full set of kinetic study has been completed for dihydrobenzoxanthones (1–4) to be competitive and reversible simple slow-binding inhibitors in  $\alpha$ -glucosidase and has rarely been reported from natural phenolic compounds ((Jenis, et. al, 2019).

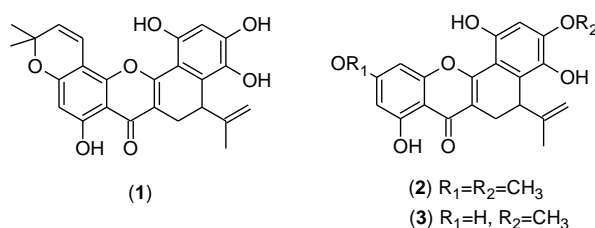


Figure 1. Dihydrobenzoxanthones

If we look carefully, it can be seen that the ring B oxidation pattern of dihydroxanthone compounds is not in accordance with the pattern of the acetate malonate pathway and the shikimat pathway which is the biogenesis pathway of the flavonoid group.

## Conclusion

Dihydrobenzoxanthone is one of flavonoid derivatives which is successfully isolated from *Artocarpus*. Dihydrobenzoxanthone is only formed from the flavone with ring B which is oxygenated with pattern of 2', 4' and 5'. the ring B oxidation pattern of dihydroxanthone compounds is not in accordance with the pattern of the acetate malonate pathway and the shikimat pathway which is the biogenesis pathway of the flavonoid group.

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## References

- Achmad, S.A., Hakim, E. H., Juliawaty, L. D., Makmur., L., & Suyatno (1996), New Prenylated Flavone from *Artocarpus champeden*, *J. Nat Prod*, **59**, 878-879. <https://doi.org/10.1021/np960330q>
- Achmad, S.A., Hakim, E. H., Juliawaty, L. D., Makmur., L., Kusuma, S. & Syah, Y. M. (1995), Eksplorasi Kimia Tumbuhan Tropis Indonesia, Seminar Nasional Etnobotani II, Yogyakarta, 112-113.
- Boonlaksiri, C., Oonanant, W., Kongsaree, P., Kittakoo, P., Tanticharoen, M., & Thebtaranonth, Y. (2000). An antimalarial stilbene from *Artocarpus integer*. *Phytochemistry*, *54*(4), 415-417. [https://doi.org/10.1016/S0031-9422\(00\)00074-1](https://doi.org/10.1016/S0031-9422(00)00074-1)
- Darmawati, A. A. S. K., Bawa, I. G. A. G., & Suirta, I. W. (2015). Isolasi dan Identifikasi Senyawa Golongan Flavonoid pada Daun Nangka (*Artocarpus heterophyllus* Lmk) dan Aktivitas Antibakteri terhadap Bakteri *Staphylococcus aureus*. *Jurnal Kimia (Journal of Chemistry)*.
- Ferlinahayati (1999). Norartokarpetin dan flavonoid terisoprenilasi dari kulit batang *Artocarpus scortechinii* King [Tesis]. Institut Teknologi Bandung. Bandung
- Hakim, A. (2010). Diversity of secondary metabolites from Genus *Artocarpus* (Moraceae). *Nusantara Bioscience*, *2*(3). <https://doi.org/10.13057/nusbiosci/n020307>
- Heyne, K., (1987), *Tumbuhan Berguna Indonesia II*, Terjemahan, Penerbit Badan Litbang Kehutanan, Jakarta, 645-646.
- Jayasinghe ULB, Samarakoon TB, Kumarihamy BMM, Hara N, & Fujimoto Y. (2008). Four new prenylated flavonoids and xanthenes from the root bark of *Artocarpus nobilis*. *Fitoterapia* *79*: 37-41. <https://doi.org/10.1016/j.fitote.2007.07.014>
- Jayasinghe, L., Balasooriya, B.A.I.S., Padmini,W.C., Hara, N., & Fujimoto Y. (2004). Geranyl chalcone derivatives with antifungal and radical scavenging properties from the leaves of *Artocarpus nobilis*, *Phytochemistry*, **65**, 1287-1290. <https://doi.org/10.1016/j.phytochem.2004.03.033>
- Jenis, J., Baiseitova, A., Yoon, S. H., Park, C., Kim, J. Y., Li, Z. P., ... & Park, K. H. (2019). Competitive  $\alpha$ -glucosidase inhibitors, dihydrobenzoxanthenes, from the barks of *Artocarpus elasticus*. *Journal of enzyme inhibition and medicinal chemistry*, *34*(1), 1623-1632. <https://doi.org/10.1080/14756366.2019.1660653>
- Khan, M.R., Omoloso, A.D., & Kihara, M., (2003) Antibacterial activity of *Artocarpus heterophyllus*, *Fitoterapia*, **74**, 501-505. [https://doi.org/10.1016/S0367-326X\(03\)00120-5](https://doi.org/10.1016/S0367-326X(03)00120-5)
- Nomura, T., Hano, S., & Aida M. (1998), *Heterocycles*, *47*(2), 1179-1205.

- <https://www.infona.pl/resource/bwmeta1.element.elsevier-6717c273-ac61-3f94-8497-dc82cb7add9b>
- Okunlola, A., & Adewusi, S. A. (2019). Development of theophylline microbeads using pregelatinized breadfruit starch (*artocarpus altilis*) as a novel co-polymer for controlled release. *Advanced pharmaceutical bulletin*, 9(1), 93. <https://dx.doi.org/10.15171%2Fapb.2019.012>
- Pratap, R., & Ram, V. J. (2014). Natural and synthetic chromenes, fused chromenes, and versatility of dihydrobenzo [h] chromenes in organic synthesis. *Chemical reviews*, 114(20), 10476-10526. <https://doi.org/10.1021/cr500075s>
- Sultanbawa, M.U.S., Surendrakumar, & Sivagnanasundram. (1989). Two pyranodihydrobenzoxanthones from *Artocarpus nobilis*. *Phytochemistry* 28(2): 599-605. [https://doi.org/10.1016/0031-9422\(89\)80059-7](https://doi.org/10.1016/0031-9422(89)80059-7)
- Syah YM, Achmad SA, Ghisalberti EL, Hakim EH, Makmur L, & Mujahidin D. (2002). Artoindonesianins Q–T, four isoprenylated flavones from *Artocarpus champeden* Spreng. (Moraceae). *Phytochem* 61 949-953. [https://doi.org/10.1016/S0031-9422\(02\)00366-7](https://doi.org/10.1016/S0031-9422(02)00366-7)
- Syah, Y.M., Juliawaty, L.D., Achmad, S.A., Hakim, E.H., & Ghisalberti, E.L, (2006) Cytotoxic prenylated flavones from *Artocarpus champeden*, *Journal Natural Medicine*, 60, 308-312. <https://link.springer.com/article/10.1007/s11418-006-0012-z>
- Wang, Y., Deng, T., Lin, L., Pan, Y., & Zheng, X. (2006). Bioassay-guided isolation of antiatherosclerotic phytochemicals from *Artocarpus altilis*. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, 20(12), 1052-1055. <https://doi.org/10.1002/ptr.1990>
- Widyawaruyanti, A., Subehan, Kalauni, S.K., Awale, S., Nindatu, M., Zaini, N.C., Syafruddin, D., Asih, P.B.S., Tezuka, Y., ... Kadota, S. (2007), New prenylated flavones from *Artocarpus champeden*, and their antimalarial activity in vitro, *Journal Natural Medicine*, 61, 410-413.
- Zhao, J., & Dixon, R. A. (2010). The 'ins' and 'outs' of flavonoid transport. *Trends in plant science*, 15(2), 72-80. <https://doi.org/10.1016/j.tplants.2009.11.006>
- Harborne, J. B. (1989). Flavonoids. In *Natural products of woody plants* (pp. 533-570). Springer, Berlin, Heidelberg. [https://link.springer.com/chapter/10.1007/978-3-642-74075-6\\_16](https://link.springer.com/chapter/10.1007/978-3-642-74075-6_16)