# OPTIMIZATION OF YELLOW PASSION FRUIT PEEL EXTRACTION METHOD USING METHANOL SOLVENT TO INCREASE ANTIOXIDANT ACTIVITY

Khoirul Ngibad\*, Muhammad Sungging Pradana, Juli Afifah, and Aulia Afiatunnisa

Medical Laboratory Technology Study Program, Faculty of Health, Universitas Maarif Hasyim Latif, Sidoarjo, East Java, Indonesia

\*Email: <u>khoirul\_ngibad@dosen.umaha.ac.id</u>

Received: November 29, 2022. Accepted: January 29, 2023. Published: January 30, 2023

**Abstract:** This study aims to determine the concentration of methanol solvent, the temperature, and the time of the shaker in the best extraction. It can produce the best antioxidant activity of the peel of the yellow passion fruit (Passiflora edulis Sims f. flavicarpa Deg) using the DPPH method. The maceration process is carried out using methanol solvents. The filtrate is concentrated with a rotary evaporator until a concentrated extract is obtained. Variations in methanol concentrations include 60, 80, and 100% (v/v) in methanol-water solvent mixtures. Shaker time variations in the extraction process include: 120, 140, and 160 minutes and shaker temperature variations in the extraction process include: 55, 65, and 75 °C. The results showed that the concentration of methanol solvent that can produce the best antioxidant activity from the peel of yellow passion fruit using the DPPH method is a concentration of 100% with an IC50 value of 31 ppm, followed by methanol solvents of 60% and 80% which produce IC<sub>50</sub> values of 35 and 41 ppm, respectively. The optimum shaker time in the extraction process is 140 minutes, and the optimum shaker temperature in the extraction process is 65 °C. The conclusion is that using 100% methanol solvent, 140 minutes shaker time, and 65 °C shaker temperature can increase antioxidant activity in vitro using the DPPH method.

Keywords: Antioxidant, Temperature, Time, Shaker, Methanol 100%, Yellow Passion Fruit.

# **INTRODUCTION**

Passion fruit is one of the plants that can thrive in tropical climates even though it is not native to Indonesia. Passion fruit has two types: yellow and purple passion fruit, each unique [1]. Many Indonesians do various processing on passion fruit, such as: making passion fruit juice [2], passion fruit syrup [3], and herbal tea [4]. The use of passion fruit is still focused on the fruit part only, while the utilization of the passion fruit Peel still needs to be widely done. According to [5], fruit peel waste has a high content of antioxidants. In addition, many studies state that passion fruit peel is effective as an antibacterial, anti-inflammatory, antihyaluronidase, anti-collagenase, antidiabetic, and antioxidant [2, 6-13].

One of the benefits and peel of passion fruit has many benefits as an antioxidant that can minimize the occurrence of oxidation reactions and prevent the formation of free radicals triggered by factors from the inside, names derived from normal metabolism, and external factors, such as drugs and pollution [14]. Antioxidants are divided into 2, namely: natural and artificial. Unlike natural antioxidants, synthetic antioxidants can provide dangerous side effects. Therefore, many researchers study to find natural sources of antioxidants [16], such as lutein, lycopene, vitamin C, vitamin E, and carotenoids that are usually found in fruits and vegetables.

Testing the antioxidant activity of the peel of purple passion fruit using extracts from ethanol solvents has been widely used in studies [6, 15-24]. But the peel of yellow passion fruit has yet to be studied much. On the other hand, passion fruit peel extracts that use methanol solvents still need to be widely used. According to [25], the antioxidant potential of passion fruit peel methanol extract is higher than that of ethanol and water extracts. Therefore, in this study, optimization was carried out in the maceration extraction process, including methanol solvent concentration, extraction time, and extraction temperature, to produce the best antioxidant activity using the DPPH method.

### **RESEARCH METHODS** Chemicals

Yellow passion fruit peel Simplicia powder was obtained east of the STKIP Muhammadiyah Kuningan Campus, Jl. Raya Cigugur RT.5 / RW.2 Kuningan, West Java. The chemicals include methanol, DPPH, mineral-free water, and ascorbic acid, purchased from Sigma Aldrich. All reagents are analytical classes.

# **Research Design**

Variations in methanol concentrations include 60, 80, and 100% (v/v) in methanol-water solvent mixtures. Shaker time variations in the extraction process include: 120, 140, and 160 minutes and shaker temperature variations in the extraction process include: 55, 65, and 75 °C.

# Procedure

#### Extraction

100 g of powder was macerated using a solvent of 1000 mL of methanol-water [26]. The maceration process is carried out for 24 hours and

then filtered into filtrate and pulp. The filtrate is concentrated with a rotary evaporator until a concentrated extract is obtained [27] [28].

## Preparation of Test Sample Solutions and Antioxidant Standards

A test sample solution of methanol extract and vitamin C was made in concentrations (20, 40, 60, 80, and 100) mg/L of 50 mL each.

# Antioxidant Test using DPPH Method

Each test sample solution and the antioxidant standard were pipetted by 0.25 mL and mixed with a  $6 \times 10$ -5 M DPPH solution of 5 mL in a test tube. Then vortex for 1 minute. After incubation for 30 minutes at room temperature, the absorbance of the reaction mixture was measured at a wavelength of 515 nm using a spectrophotometer to obtain the absorbance value of the sample. A blank solution consisting of methanol in DPPH solution is prepared and measured at the same wavelength to obtain the control absorbance value. The experiment was carried out with three repetitions. Antioxidant activity is calculated using equation (2).

Antioxidant activity (%) = (control absorbance – sample absorbance)/control absorbance  $\times$  100 %

Next, a graph of the relationship between the sample concentration (x-axis) and the percent radical inhibition of DPPH (y-axis) was created. The calculation of the  $IC_{50}$  value based on the formula of the linear regression equation

# **RESULTS AND DISCUSSION**

# The yield of Concentrated Extract of Yellow Passion Fruit Peel

In this study, optimization was carried out in the extraction process by maceration of yellow passion fruit peel with the yield of % amendment shown in Table 1. Based on the table, the result can be obtained that the greater the concentration of methanol solvent used in the maceration process, the greater the % yield produced. 100% methanol solvent is the best solvent to extract the content of secondary metabolite compounds from the peel of yellow passion fruit with a % amendment of 13.26%. The results showed that the level of the polarity of the secondary metabolite compounds from the peel of the yellow passion fruit corresponds to the polarity of the 100% methanol solvent so that it can maximize % the yield.

Variations of methanol solvents	Simplicia weight (g)	Concentrated extract weight (g)	%Yield
Effect of methanol solvent concer	ntration		
Methanol 60%*	100	12.06	12.06
Methanol 80%**	100	12.88	12.88
Methanol 100%***	100	13.26	13.26
Effect of shaker time in the extract	tion process		
120 minutes <sup>1</sup>	100	6.51	6.51
140 minutes <sup>2</sup>	100	7.08	7.08
160 minutes <sup>3</sup>	100	7.34	7.34
Effect of shaker temperature in ex	traction process		
55°C <sup>a</sup>	100	6.06	6.06
$65^{\circ}C^{b}$	100	6.58	6.58
75°C°	100	7.04	7.04

Table 1. The yield of concentrated extracts of yellow passion fruit peel from various treatments

Information

\* : Yellow passion fruit peel extracted with 60% methanol solvent

\*\* : Yellow passion fruit peel extracted with 80% methanol solvent

\*\*\* : Yellow passion fruit peel extracted with 100% methanol solvent

<sup>1</sup> : Extracted yellow passion fruit peel with complaint time for 120 minutes

<sup>2</sup> : Extracted yellow passion fruit peel with complaint time for 140 minutes

<sup>3</sup> : Extracted yellow passion fruit peel with complaint time for 160 minutes

<sup>a</sup> : Yellow passion fruit peel extracted at a shaker temperature of 55°C in the extraction process

<sup>b</sup> : Yellow passion fruit peel extracted at a shaker temperature of 65°C in the extraction process

<sup>c</sup> : Yellow passion fruit peel extracted at a shaker temperature of 75°C in the extraction process

In addition, to the influence of methanol solvent concentration, the influence of shaker time in the extraction process was also studied. The results showed that using shakers in the maceration process for 160 minutes could increase % yield. The use of shakers in this study increased the contact of methanol solvents with samples of yellow passion fruit powder [29]. Furthermore, the temperature of J. Pijar MIPA, Vol. 18 No. 1, January 2023: 77-83 DOI: 10.29303/jpm.v18i1.4434 ISSN 1907-1744 (Cetak) ISSN 2460-1500 (Online)

the shaker in the extraction process also affects the % amendment. The greater the temperature of the shaker, the greater the % of the resulting amendment. In the study, the temperature of the shaker that produced the highest % yield was 75°C. The higher temperature of the shaker used in the maceration process can accelerate the contact between the sample and the solvent [30].

# Antioxidant Activity of Concentrated Extract of Yellow Passion Fruit Peel

The relationship between the concentration of methanol extract from the yellow passion fruit peel and the percentage of DPPH radical inhibition in the treatment of various methanol solvent concentrations is shown in Figure 1. The effect of variations in methanol solvent concentrations on antioxidant activity using the DPPH method was studied with the results shown in Table 2. The smaller IC value of 50 indicates that the extract has better antioxidant activity [31]. Based on the table, it can be seen that the IC value of 50 decreases further following an increase in the concentration of methanol solvent used in the extraction process by maceration. Methanol solvents with a concentration of 100% or in the absence of additional water solvents in this study were able to produce the best IC 50 value of 31 ppm, followed by methanol solvents with a concentration of 60% and methanol solvents with a concentration of 80% which produced IC 50 values of 35 and 41 ppm, respectively.

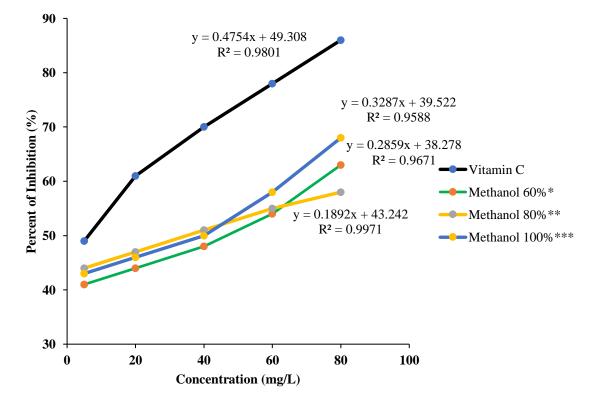


Figure 1. The relationship between the concentration of methanol extract of yellow passion fruit peel and the percentage of DPPH radical inhibition in the treatment of various methanol solvent concentrations

Table 2. Effect of methanol solvent concentration on antioxidant activity of yellow passion fruit peel extract

Variations in methanol solvent concentrations	Linear line equation	Linearity	IC <sub>50</sub> value (ppm)
Vitamin C	y = 0.4754x + 49,308	$R^2 = 0.9801$	1.46
Methanol 60%*	y = 0.2859x + 38,278	$R^2 = 0.9671$	41
Methanol 80% **	y = 0.1892x + 43,242	$R^2 = 0.9971$	35
Methanol 100% ***	y = 0.3287x + 39,522	$R^2 = 0.9588$	31

Information

\* : Yellow passion fruit peel extracted with 60% methanol solvent

\*\* : Yellow passion fruit peel extracted with 80% methanol solvent

\*\*\* : Yellow passion fruit peel extracted with 100% methanol solvent

The relationship between the concentration of methanol extract of yellow passion fruit peel and the percentage of DPPH radical inhibition in the treatment of shaker time variation in the maceration extraction process is shown in Figure 2. The effect of shaker time variation in the maceration extraction process on antioxidant activity using the DPPH method was also studied, with the results shown in Table 3. Based on the table, it can be known that the time of the shaker in the best extraction process that produces the best antioxidant activity is 140 minutes. On the other hand, a shaker time of 160 minutes can decrease antioxidant activity using the DPPH method. It can be caused by the length of time the shaker can damage the content of secondary metabolite compounds contained in the peel of yellow passion fruit and potentially increase the process of loss of secondary metabolite compounds in the extracted solution due to evaporation [32].

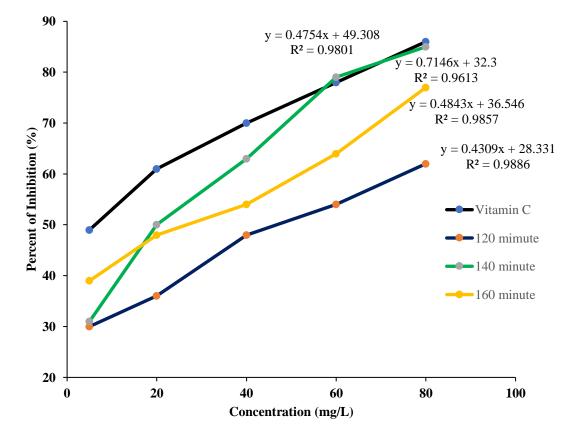


Figure 2. The relationship between the concentration of methanol extract of yellow passion fruit peel and the percentage of DPPH radical inhibition in the treatment of shaker time variation in the maceration extraction process

Table 3 Effect of shaker time in the extraction process o	n the antioxidant activity of yellow passion fruit peel

Shalan tinta mariatiana	extract	T in conita-	IC
Shaker time variations	Linear line equation	Linearity	IC <sub>50</sub> value (ppm)
Vitamin C	y = 0.4754x + 49.308	$R^2 = 0.9801$	1.46
120 minutes <sup>1</sup>	y = 0.4309x + 28.331	$R^2 = 0.9886$	50.29
140 minutes <sup>2</sup>	y = 0.7146x + 32.3	$R^2 = 0.9613$	24.77
160 minutes <sup>3</sup>	y = 0.4843x + 36.546	$R^2 = 0.9857$	27.78

Information:

<sup>1</sup>: Extracted yellow passion fruit peel with complaint time for 60 minutes

<sup>2</sup>: Extracted yellow passion fruit peel with complaint time for 80 minutes

<sup>3</sup>: Extracted yellow passion fruit peel with complaint time for 100 minutes

Furthermore, the relationship between the concentration of methanol extract of yellow passion fruit peel and the percentage of DPPH radical inhibition in the treatment of shaker temperature variations in the maceration extraction process is shown in Figure 3. The effect of shaker temperature variations in the maceration extraction process on antioxidant activity using the DPPH method was also studied, with the results shown in **Table 4.** Based on the table, it can be known that the

temperature of the shaker in the best extraction process that produces the best antioxidant activity is 65 °C. On the other hand, the temperature of the shaker set at 75 °C can cause a decrease in antioxidant activity using the DPPH method. The high temperature of the shaker can damage the content of secondary metabolite compounds contained in the peel of yellow passion fruit and potentially increase the loss of secondary metabolite compounds in the extracted solution due to evaporation [32].

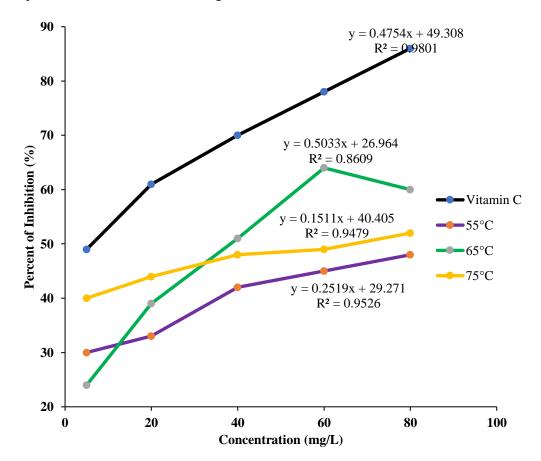


Figure 2. The relationship between the concentration of methanol extract of yellow passion fruit peel and the percentage of DPPH radical inhibition in the treatment of shaker temperature variations in the maceration extraction process

Table 4. Effect of shaker temperature in the extraction process on the antioxidant activity of yellow passion fruit peel extract

	peerextract		
Shaker temperature variations	Linear line equation	Linearity	IC <sub>50</sub> value (ppm)
Vitamin C	y = 0.4754x + 49.308	$R^2 = 0.9801$	1.46
$55^{\circ}C^{a}$	y = 0.2519x + 29.271	$R^2 = 0.9526$	82
65°C <sup>b</sup>	y = 0.5033x + 26.964	$R^2 = 0.8609$	45.77
75°C°	y = 0.1511x + 40.405	$R^2 = 0.9479$	63.5

Information:

<sup>a</sup>: Yellow passion fruit peel extracted at a shaker temperature of 55°C in the extraction process

<sup>b</sup>: Yellow passion fruit peel extracted at a shaker temperature of 65°C in the extraction process

<sup>c</sup>: Yellow passion fruit peel extracted at a shaker temperature of 75°C in the extraction process

## CONCLUSION

The concentration of methanol solvent that can produce the best antioxidant activity from the peel of yellow passion fruit using the DPPH method is a concentration of 100% with an IC50 value of 31 ppm, followed by methanol solvents of 60% and 80%, which produce IC<sub>50</sub> values of 35 and 41 ppm, respectively. The optimum shaker time in the extraction process is 140 minutes, and the optimum shaker temperature in the extraction process is 65 °C. The conclusion is that using 100% methanol solvent, 140 minutes shaker time, and 65 °C shaker temperature can increase antioxidant activity in vitro using the DPPH method.

## ACKNOWLEDGEMENTS

Thanks are conveyed to the Academic Directorate of Vocational Higher Education Directorate General of Vocational Education Ministry of Education, Culture, Research, And Technology for Fiscal Year 2022 which has funded this Penelitian Dosen Pemula (PDP) with research contract Number: 008/SP2H/PPKM/LL7/2022 Dated June 23, 2022 and 040/B-02/LPPM/UMAHA/VII/2022 Dated July 12, 2022.

## REFERENCES

- Munda, M. Perbandingan Daya Antioksidan Sari Buah Markisa Ungu (Passiflora edulis F. Edulis Sims) dengan Sari Buah Markisa Kuning (P. edulis Sims F. Flavicarpa Deg) Menggunakan Metode DPPH.
- [2] Anabel, A., Wijaya, C. D., & Lokanata, S. (2020). Uji Efektivitas Antibakteri Ekstrak Kulit Buah Markisa Ungu (Passiflora edulis sims) terhadap Staphylococcus aureus. Healthy Tadulako Journal (Jurnal Kesehatan Tadulako), 6(3), 79-85.
- [3] Kusumah, S. H., Pebrianti, S. A., & Maryatilah, L. (2021). Uji Aktivitas Antioksidan Buah dan Sirup Markisa Ungu Menggunakan Metode DPPH. Jurnal Fakultas Teknik Kuningan, 2(1), 25-32.
- [4] Permana, A. D. (2021). Analisa Fitokimia dan Aktivitas Antioksidan Serbuk Teh Hitam Buah Markisa Kuning. Univ. Sriwij.
- [5] Musika, S., Pokratok, N., Pliankratoke, J., Khongla, C., Kupradit, C., Ranok, A., & Mangkalanan, S. (2021). Antioxidant, Antityrosinase and Antibacterial Activities of Fruit Peel Extracts. Int. J. Agricult. Technol, 17, 1447-1460.
- [6] Cazarin, C. B. B., Rodriguez-Nogales, A., Algieri, F., Utrilla, M. P., Rodríguez-Cabezas, M. E., Garrido-Mesa, J. & Gálvez, J. (2016). Intestinal Anti-inflammatory Effects of Passiflora edulis Peel in the Dextran Sodium Sulphate Model of Mouse Colitis. Journal of Functional Foods, 26, 565-576.
- [7] Hartanto, S., Lister, I. N. E., & Fachrial, E. (2019). A Comparative Study of Peel and Seed Extract of Passion Fruit (Passiflora edulis) as Anti Collagenase. Am. Sci. Res. J. Eng. Technol. Sci, 54, 42-48.
- [8] Khoirot, U. (2019). Efek Pemberian Rebusan Kulit Markisa Ungu (Passiflora edulis) Sebagai Antidiabetik Terhadap Gambaran Histopatologi Pankreas Tikus Yang Diinduksi Streptozotosin (Doctoral dissertation).
- [9] Wong, Y. S., Sia, C. M., Eng, H., Ang, Y. K., Chang, S. K., & Yim, H. S. (2014). Influence of Extraction Conditions on Antioxidant Properties of Passion Fruit (Passiflora edulis)

Peel. Acta Scientiarum Polonorum Technologia Alimentaria, 13(3), 257-265.

- [10] Vera, K., Raif, A., & Ikhtiari, R. (2019). Antioxidant and Anti-elastase Activity of Seed and Peel Extract of P. edulis. Am. Sci. Res. J. Eng. Technol. Sci, 53, 43-48.
- [11] Reveny, J., & Ginting, H. (2018). Antioxidant Test, Phenolic And Flavonoid Content Ethanol Extract And Ethyl Acetate Fraction Of Purple Passion Fruit Peel (Passiflora edulis Sims.).
- [12] Ramli, A. N. M., Manap, N. W. A., Bhuyar, P., & Azelee, N. I. W. (2020). Passion Fruit (Passiflora edulis) Peel Powder Extract and Its Application towards Antibacterial and Antioxidant Activity on the Preserved Meat Products. SN Applied Sciences, 2, 1-11.
- [13] Armin, F., Ermadanis, E., & Rasyid, R. (2017). Analisis Senyawa Fenolat dan Uji Aktivitas Antioksidan Buah Markisa (Passiflora edulis Sims) Secara Spektrofotometri Visibel. Jurnal Farmasi Higea, 6(2), 117-125.
- [14] Ngibad, K., & Lestari, L. P. (2020). Aktivitas Antioksidan dan Kandungan Fenolik Total Daun Zodia (Evodia suaveolens). ALCHEMY Jurnal Penelitian Kimia, 16(1), 94-109.
- [15] Ginting, H., Dalimunthe, A., & Pratama, E. K. (2018, October). Kajian Ketoksikan Ekstrak Etanol Kulit Buah Markisah Ungu (Passiflora edulis Sims.) Terhadap Hati Mencit. In Talenta Conference Series: Tropical Medicine (TM) (Vol. 1, No. 1, pp. 257-263).
- [16] Septiani, N. (2018). Uji Aktivitas Antibakteri Ekstrak Etanol dan Fraksi-Fraksi Kulit Buah Markisa Ungu (Passiflora edulis Sims) terhadap Bakteri Staphylococcus epidermidis di Laboratorium Fitokimia dan Mikrobiologi Farmasi USU (Doctoral dissertation, Universitas Sumatera Utara).
- [17] Nazliniwaty, N., Harun, F. R., Putra, E. D. L., & Nerdy, N. (2020). Antiaging Activity of Gel Preparation Containing Three Varieties of Passion Fruit Peel Ethanolic Extract. Open Access Macedonian Journal of Medical Sciences, 8(A), 170-174
- [18] Durling, N. E., Catchpole, O. J., Grey, J. B., Webby, R. F., Mitchell, K. A., Foo, L. Y., & Perry, N. B. (2007). Extraction of Phenolics and Essential Oil From Dried Sage (Salvia officinalis) Using Ethanol–Water Mixtures. Food chemistry, 101(4), 1417-1424.
- [19] Zhang, Z. S., Li, D., Wang, L. J., Ozkan, N., Chen, X. D., Mao, Z. H., & Yang, H. Z. (2007). Optimization of Ethanol–Water Extraction of Lignans From Flaxseed. Separation and Purification Technology, 57(1), 17-24.
- [20] Kandandapani, S., Balaraman, A. K., & Ahamed, H. N. (2015). Extracts of Passion Fruit Peel and Seed of Passiflora edulis (Passifloraceae) Attenuate Oxidative Stress in

Diabetic Rats. Chinese journal of natural medicines, 13(9), 680-686

- [21] dos Santos, E. K. R., Azoubel, P. M., & Gouveia, E. R. (2017). Better Pectin Yield from Passion Fruit Peel (Passiflora edulis f. flavicarpa): From Shaker or Ultrasound? A comparison. Waste and biomass valorization, 8, 905-910.
- [22] Herrera-Ramirez, J., Meneses-Marentes, N., & Tarazona Díaz, M. P. (2020). Optimizing the Extraction of Anthocyanins From Purple Passion Fruit Peel Using Response Surface Methodology. Journal of Food Measurement and Characterization, 14(1), 185-193.
- [23] Xiong, F., Li, X., Zheng, L., Hu, N., Cui, M., & Li, H. (2019). Characterization and Antioxidant Activities of Polysaccharides from Passiflora edulis Sims Peel Under Different Degradation Methods. Carbohydrate polymers, 218, 46-52.
- [24] Musika, S., Pokratok, N., Pliankratoke, J., Khongla, C., Kupradit, C., Ranok, A., & Mangkalanan, S. (2021). Antioxidant, Antityrosinase and Antibacterial Activities of Fruit Peel Extracts. Int. J. Agricult. Technol, 17, 1447-1460.
- [25] Cazarin, C. B. B., da Silva, J. K., Colomeu, T. C., de Lima Zollner, R., & Marostica Jr, M. R. (2014). Antioxidant Capacity and Chemical Composition of Passion Fruit Peel (Passiflora edulis). Ciência Rural, 44(9), 1769-1775.
- [26] Handayani, H., Sriherfyna, F. H., & Yunianta, Y. (2016). Ekstraksi Antioksidan Daun Sirsak Metode Ultrasonic Bath (Kajian Rasio Bahan: Pelarut dan Lama Ekstraksi)[In Press Januari 2016]. Jurnal Pangan dan Agroindustri, 4(1).
- [27] Ngibad, K. (2019). Phytochemical Screening of Sunflower Leaf (Helianthus annuus) and Anting-Anting (Acalypha indica Linn) Plant Ethanol Extract. Borneo Journal of Pharmacy, 2(1), 24-30.
- [28] Ngibad, K. (2019). Efektivitas Kombinasi Ekstrak Etanol Daun Bunga Matahari dan Tanaman Anting-Anting sebagai Antimalaria Secara In Vivo. Jurnal Farmasi Galenika (Galenika Journal of Pharmacy)(e-Journal), 5(1), 12-19.
- [29] Arfisa, C., Widiyana, A. P., and Bintari, Y. R. (2020). Perbandingan Kadar Merkuri (Hg) dan Aktivitas Antioksidan Ekstrak Metanolik Akar Eceng Gondok (Eichornia crassipes) di Daerah Lawang dan Pasuruan. J. Kedokt. Komunitas, 8(21), 1–8
- [30] Alfajri, S., Agustina, F., Sari, N. P., & Pramuanggit, P. N. (2018). Uji Resistensi Bakteri Vibrio parahaemolyticus terhadap Ekstrak Makroalga Halimeda discoidea,

Halymenia dilatata dan Dictyota dichotoma. Simbiosa, 7(1), 33-46.

- [31] Jusmiati, J., Rusli, R., & Rijai, L. (2015). Aktivitas antioksidan Kulit Buah Kakao Masak dan Kulit Buah Kako Muda. Jurnal Sains dan Kesehatan, 1(1), 34-39.
- [32] Kemit, N., Widarta, I. W. R., & Nocianitri, K. A. (2017). Pengaruh Jenis Pelarut dan Waktu Maserasi Terhadap Kandungan Senyawa Flavonoid dan Aktivitas Antioksidan Ekstrak Daun Alpukat (Persea americana Mill). Jurnal Ilmu Dan Teknologi Pangan (Itepa) Universitas Udayana.